

FROM SCREENING TO PRECISION CARE: VALIDATING BORDERLINE *LDLR* VARIANTS THROUGH FLOW CYTOMETRY

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BACKGROUND

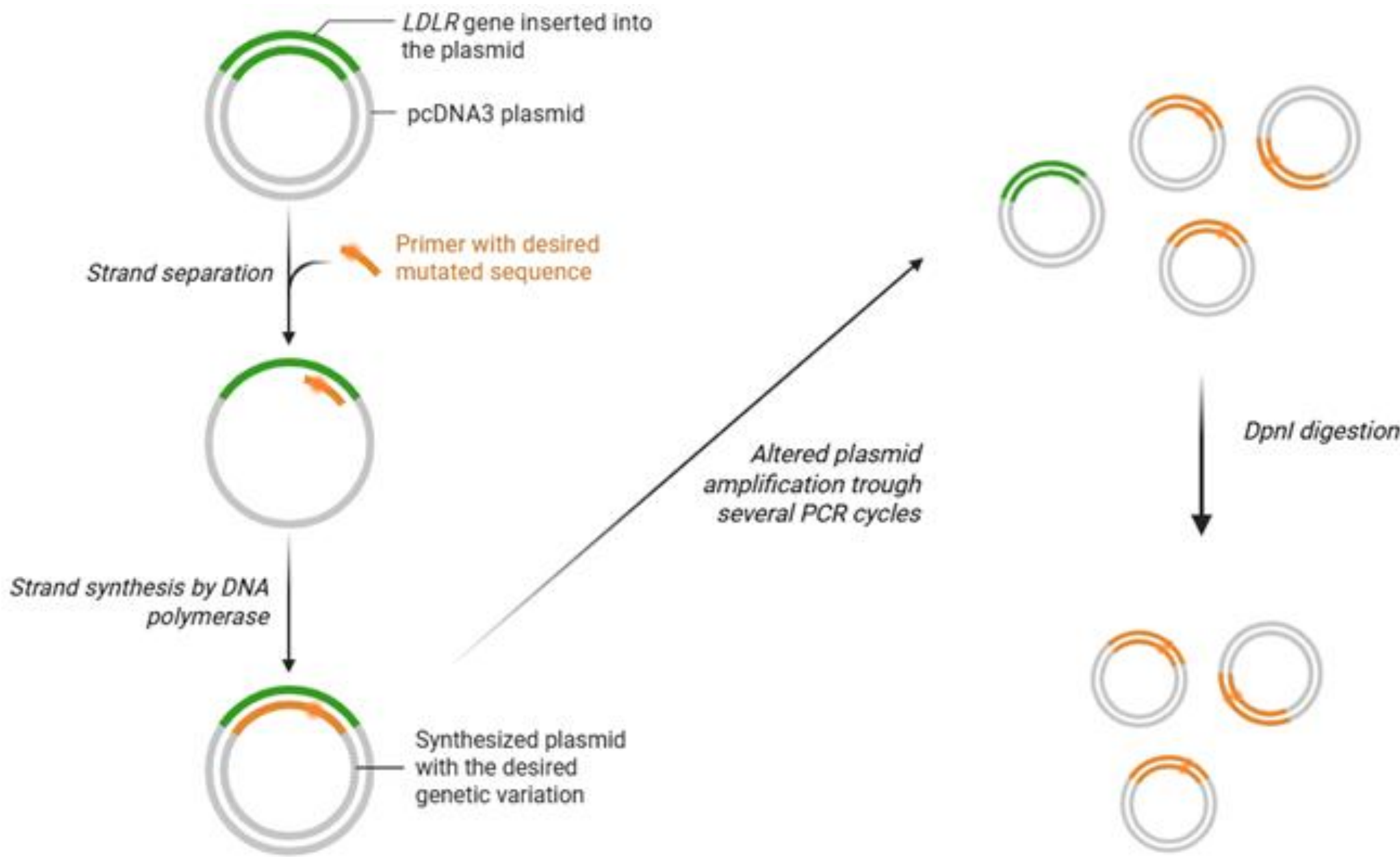
- Familial Hypercholesterolaemia (FH) is a semidominant disorder primarily caused by variants in *LDLR*, leading to elevated LDL-cholesterol levels. Functional characterisation is essential for accurate diagnosis and clinical management.
- High-throughput assays start to emerge as screening tools (e.g., PerMedFH), but they provide an overall measure of LDLR activity and may mask defects in specific steps of the LDLR cycle.
- Variants with borderline activity (70–90%) remain particularly challenging, as current FH VCEP guidelines do not consider these results as definitive functional evidence.

AIM

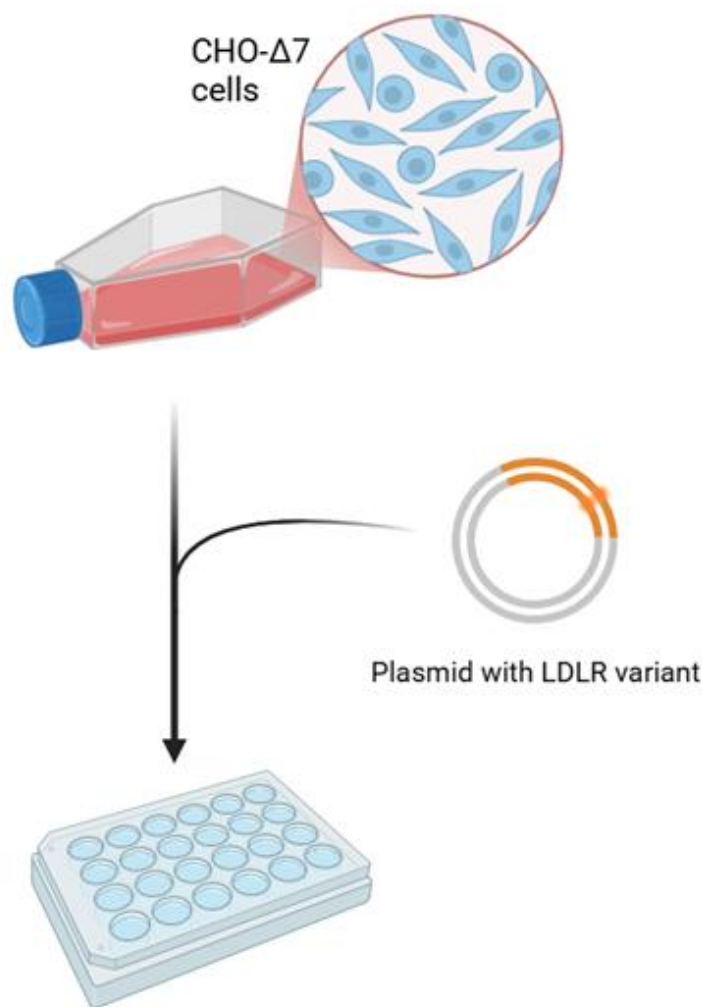
To determine whether LDLR variants with borderline activity (70–90%) exhibit functional defects when assessed by flow cytometry.

METHODS

1 Site-directed Mutagenesis

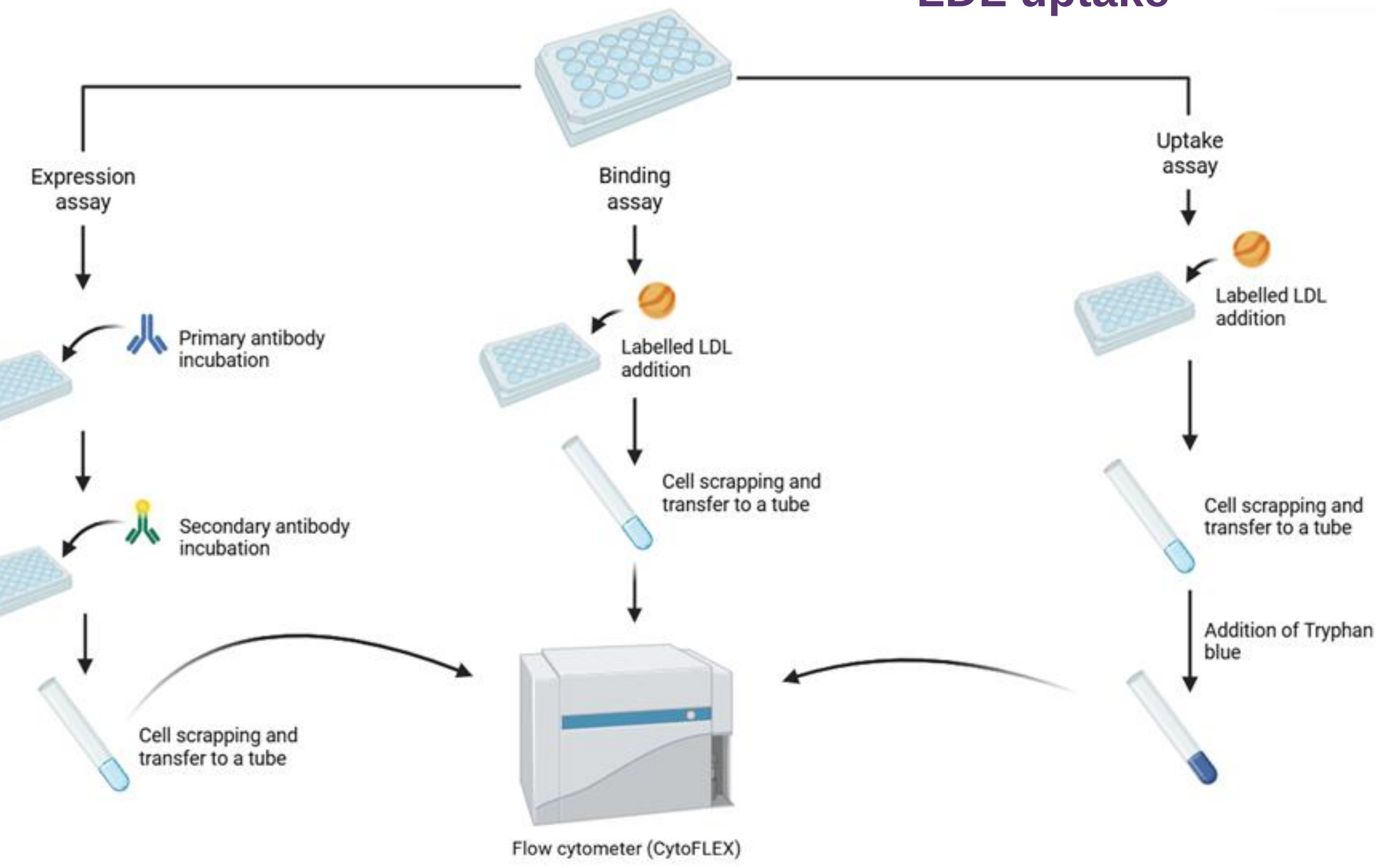


2 Transfection into CHO-IΔ7 Cells



3 Flow Cytometry Assays:

- LDLR expression
- LDL binding
- LDL uptake



RESULTS

- Most variants (15/16) showed normal LDLR expression, binding, and uptake (Figure 1), meaning that these variants don't affect the LDL metabolism.
- Variant **c.1190C>T/p.(Ser397Phe)** showed:
 - Normal expression
 - Normal binding
 - Impaired LDL uptake (<5%)
 - This variant is responsible for an increase in LDL cholesterol levels.
- Flow cytometry revealed functional defects not detected by high-throughput screening.
- Functional classification may differ depending on the assay used, particularly for variants with borderline activity (70–90%).
- p.(Ser397Phe) shows a selective defect in LDL uptake (<70%), supporting pathogenic classification (FH VCEP).
- Even in the absence of detectable defects, flow cytometry provides high-level evidence that strengthens variant classification (FH VCEP).

Flow cytometry reveals a functional defect in p.(Ser397Phe) not detected by screening assays

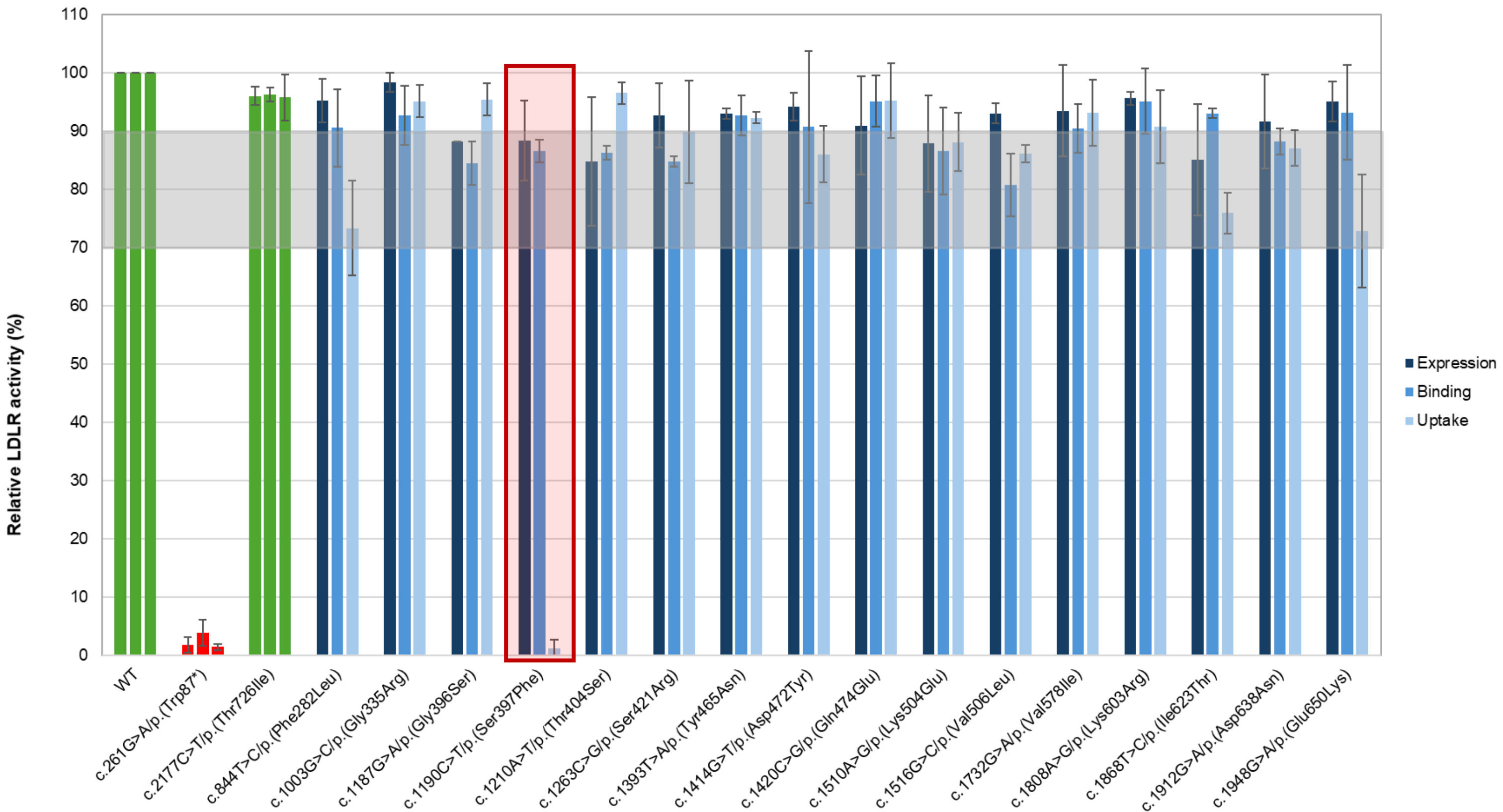


Figure 1. Results of flow cytometry from the three functional assays performed for 16 *LDLR* variants. First column (dark blue) represents LDLR expression after incubation with primary antibody (anti-LDL receptor mouse monoclonal, IgG-C7) and secondary antibody (Alexa Fluor™ 488 goat anti-mouse IgG). Second column (medium blue) represents LDL receptor's binding capacity after 3h incubation at 4°C with FITC-labelled LDL. Third column (light blue) represents LDL receptor's uptake capacity after 3h incubation at 37°C with FITC-labelled LDL and treatment with Trypan Blue. Variant p.(Trp87*) (red) represents a defective control and variant p.(Thr726Ile) (green) represents a normal control.

CONCLUSIONS

- Flow cytometry enables comprehensive assessment of LDLR function
- Identifies hidden defects in variants with borderline activity
- Strengthens variant classification under FH VCEP guidelines
- Provides a critical validation step following high-throughput screening
- Supports more accurate and personalised FH diagnosis

IMPACT

Integrating targeted functional assays into screening pipelines improves diagnostic accuracy and helps refine clinical decision-making in FH.